

Furan formation during storage and reheating of sterilized vegetable purées

Stijn Palmers**, Tara Grauwet, Carolien Buvé, Lore Van de Vondel, Biniam T. Kebede,
Marc E. Hendrickx, Ann Van Loey*

All authors are affiliated to:

Laboratory of Food Technology

Leuven Food Science and Nutrition Research Center (LFoRCe)

Department of Microbial and Molecular Systems (M²S)

KU Leuven

Kasteelpark Arenberg 22 box 2457

B-3001 Heverlee

Belgium

<http://www.biw.kuleuven.be/m2s/clmt/lmt>

*Author to whom correspondence should be addressed

Fax: +32-16-32.19.60

Telephone: +32-16-32.15.67

E-mail: ann.vanloey@biw.kuleuven.be

**Author to whom correspondence should be addressed during submission process

Fax: +32-16-32.19.60

Telephone: +32-16-37.95.22

E-mail: stijn.palmers@biw.kuleuven.be

26 **Furan formation during storage and reheating of sterilized vegetable purées**

27

28 To this day, research for furan mitigation has mostly targeted the levels of food production and handling
29 of prepared foods by the consumer. However, part of the furan concentrations found in commercially
30 available food products might originate from chemical deterioration reactions during storage. A range of
31 individual vegetable purées was stored at two different temperatures to investigate the effects of storage
32 on the furan concentrations of shelf-stable, vegetable-based foods. After 5 months of storage at 35 °C
33 (temperature-abuse conditions), a general increase in furan concentrations was observed. The furan
34 formation during storage could be reduced by storing the vegetable purées at a refrigerated temperature
35 of 4 °C, at which the furan concentrations remained approximately constant for at least 5 months.
36 Following storage, the vegetable purées were briefly reheated to 90 °C, to simulate the effect of the final
37 preparation step before consumption. As contrary to storage, furan concentrations decreased as a result
38 of evaporative losses. Both refrigerated storage and the reheating step prior to consumption showed the
39 potential of mitigation measures for furan formation in vegetable-based foods (e.g. canned vegetables,
40 ready-to-eat soups, sauces or baby foods). Next to furan, the vegetable purées were analyzed for 2- and
41 3-methylfuran. Tomato was very susceptible to the formation of both alkylated derivatives of furan, as
42 opposed to the other vegetables in this study. Methylfuran concentrations rapidly decreased during
43 storage, which was contrary to the results observed for furan.

44

45 Keywords: furan; methylfuran; vegetable; sterilization; storage; reheating.

46 Word count: 4706 (main text)

47

48

49

50

51

52 **Introduction**

53 Furan (C_4H_4O) is a small organic molecule with high volatility. In 1995, furan was classified as
54 ‘possibly carcinogenic’ to humans after it was proven to be carcinogenic in rats and mice (International
55 Agency for Research on Cancer (IARC) 1995). Furan can be formed in a variety of heat-treated foods.
56 The highest concentrations are found in coffee products and canned or jarred foods, products that are
57 subjected to an intensive heat treatment for roasting or sterilization purposes (US Food and Drug
58 Administration (FDA) 2009; European Food Safety Authority (EFSA) 2011). In the literature, many
59 ways leading to the formation of furan have been reported, the major precursors being sugars (alone or
60 in combination with amino acids), ascorbic acid and unsaturated fatty acids, followed by amino acids
61 and carotenoids (Locas & Yaylayan 2004; Becalski & Seaman 2005; Fan 2005; Mark et al. 2006;
62 Limacher et al. 2007; Limacher et al. 2008; Owczarek-Fendor et al. 2010; Owczarek-Fendor et al. 2011;
63 Van Lancker et al. 2011; Huang et al. 2011; Owczarek-Fendor et al. 2012). Vegetable-based foods (e.g.
64 canned vegetables, ready-to-eat soups, sauces or baby foods) are particularly susceptible to furan
65 formation, because they are natural mixtures of all known furan precursors, combined with a low acidity
66 that implies a sterilization treatment to obtain shelf-stable products.

67 A recent risk evaluation by the Joint FAO/WHO Expert Committee on Food Additives (2011) has
68 indicated a human health concern for furan and consequently, actions should be taken to minimize
69 exposure to an acceptable level. The most obvious target for furan mitigation is the thermal processing
70 step for preservation, e.g. by optimization of the conventional heating process or application of an
71 alternative processing technique. In this context, high-pressure high-temperature (HPHT) processing has
72 recently presented itself as an interesting alternative for furan reduction in vegetable-based foods
73 (Palmers et al. 2014). Following a HPHT treatment (600 MPa, 117 °C, $F_0 = 5$ min), the furan
74 concentrations of a wide range of vegetable purées decreased to levels close to the analytical limits (1-2
75 ng/g purée). At this moment, the specific applications of HPHT processing are still under research, and
76 equipment and operating costs might limit its use to high value-added foods. For many other products,
77 conventional thermal processing would remain the standard technique of sterilization. For these

78 products, mitigation strategies aiming at an intervention in the reaction mechanism of furan formation
79 (e.g. by changing the product composition) might have the best potential for furan reduction. As a first
80 step towards product optimization of vegetable-based foods, vitamin C and sugars were identified as the
81 major precursors and possible targets for furan mitigation (Palmer et al. 2014). In any way, the
82 reduction of furan concentrations in thermally treated foods has proven to be challenging, both because
83 of the microbial safety standards to comply with, and the variety of possible furan precursors. Therefore,
84 researchers have been looking for other, complementary ways to reduce furan concentrations. Some
85 researchers have targeted the consumer level, however also with varying success. Up to 50% of the
86 initial furan concentrations could be lost by reheating and/or stirring of the products (Zoller et al. 2007;
87 Roberts et al. 2008; Kim et al. 2009). Nevertheless, all consumer advice is considered premature at this
88 point, because the reduction in furan concentrations was not consistently observed in other studies
89 (Hasnip et al. 2006; Lachenmeier et al. 2009).

90 Following sterilization, most vegetable-based food products are considered to be microbiologically
91 stable, and hence can be stored for a long time (~years) at temperatures exceeding room temperature.
92 However, regardless of the product or the applied intensity of the preservation process, there is always a
93 certain potential left for chemical changes during subsequent storage. This chemical instability of heat-
94 treated foods seems to be ignored in the literature concerning furan as a process-induced contaminant.
95 Theoretically, storage can result in both an increase (due to enhanced degradation of its precursors and
96 reaction products) or a decrease of the furan concentration (due to degradation reactions of furan itself),
97 which is important information in the context of furan mitigation. The product stability depends on
98 numerous intrinsic (e.g. water activity, pH) and extrinsic parameters (e.g. temperature, time, light)
99 (Narayan 1997), making it difficult to predict whether the changes in product composition during
100 storage will have an observable effect on the level of furan concentration. Regarding vegetable-based
101 foods, major furan precursors such as vitamin C and sugars are known to be degraded during storage
102 (Taub & Singh 1997). As a result, these products might be identified as possible risk matrices with
103 regard to furan formation during storage. Therefore, the objective of the present study was to gain

104 insight into the effects of storage and reheating on the furan concentrations of sterilized, vegetable-
105 based foods. As recently proposed by Becalski et al. (2010) and adopted in the recommendations of the
106 Joint FAO/WHO Expert Committee on Food Additives (2011), also the concentrations of 2- and 3-
107 methylfuran were monitored. Both alkylated derivatives of furan might be of toxicological interest,
108 since animal studies have shown that they can be metabolically activated in a similar way as furan (Gill
109 et al. 2014a; Gill et al. 2014b).

110

111 **Material and methods**

112 ***Preparation of the vegetable purées***

113 Seven vegetables were freshly bought at a local supplier. The selection included broccoli (*Brassica*
114 *oleracea* Italica Group), orange carrot (*Daucus carota* ‘Nerac’), pea (*Pisum sativum*, variety unknown),
115 potato (*Solanum tuberosum* ‘Artemis’), pumpkin (*Cucurbita maxima* ‘Hokkaido’), tomato (*Solanum*
116 *lycopersicum* ‘CLX 38197’) and red beet (*Beta vulgaris* ‘Pablo’). All vegetables are commonly used
117 ingredients of ready-to-eat foods, such as jarred baby foods, soups and sauces, products found to be
118 susceptible to furan formation in recent monitoring studies (US Food and Drug Administration (FDA)
119 2009; European Food Safety Authority (EFSA) 2011). Furthermore, the selected vegetables differ in
120 their composition regarding furan precursors. The vegetables were carefully washed and cut into
121 standardized pieces of approximately 1 cm thickness, before vacuum-packing in low-density
122 polyethylene bags. To assure that all the changes observed during storage or reheating were chemical,
123 the vegetable pieces were blanched at 95 °C for 8 min in a water bath (WBU 45, Memmert, Schwabach,
124 Germany). The blanching conditions were validated using a qualitative and quantitative peroxidase test
125 (Adebooye et al. 2008; Vervoort et al. 2012). After blanching, the plastic bags were immediately cooled
126 in iced water for 10 min, frozen in liquid nitrogen, and stored in a freezer at -40 °C. Prior to purée
127 preparation, the vegetable pieces were thawed overnight in a cold room at 4 °C. The vegetable pieces
128 were mixed with a standardized amount of deionized water and blended for 1 min to obtain seven
129 individual vegetable purées. Finally, the vegetable purées were subjected to a high pressure

130 homogenization (GEA Niro Soavi, Parma, Italy) at 100 MPa, to ensure the stability of the purées during
131 the thermal treatment for sterilization and subsequent storage of the samples.

132

133 ***Thermal sterilization***

134 The individual vegetable purées were subjected to a thermal sterilization in a static Steriflow pilot
135 retort (Barriquand, Roanne, France). Due to their inert nature, glass jars (100 ml volume, 95 mm height
136 and 45 mm diameter) were used as sample holders. The jars were filled with 85 ± 0.5 g of vegetable
137 purée and then closed with metal lids. Next, they were loaded into the retort and sterilized at a retort
138 temperature of 117 °C. The holding time (± 30 min) was calculated in advance to obtain an industrially
139 relevant process value $F_{121.1}^{10\text{ }^{\circ}\text{C}}(F_0) = 5$ min in the coldest point of the product. Temperature profiles in
140 the retort and in the product were recorded using type T thermocouples (Ellab, Hilleroed, Denmark)
141 (results not shown). After sterilization, the glass jars were immediately transferred to iced water to
142 reduce further chemical reactions. A sample was taken for (methyl)furan analysis, and the remaining
143 jars were used for storage and reheating experiments.

144

145 ***Storage conditions***

146 After sterilization, the vegetable purées were placed in incubators for storage at constant temperature,
147 and protected from light. To select appropriate storage conditions, the principles of accelerated shelf life
148 testing (Toledo 2007) were applied. From a worst-case perspective, part of the sterilized vegetable
149 purées were stored at 35 °C for 5 months. Based on a theoretical temperature coefficient of (Q_{10}) of 2,
150 these storage conditions should correspond to a storage time of approximately one year at 20 °C. To
151 evaluate the potential of storage at optimal (refrigerated) storage conditions, the other part of the
152 vegetable purées were stored at 4 °C for 5 months. After storage, the glass jars were again transferred to
153 iced water to reduce further chemical reactions. A sample was taken for (methyl)furan analysis, and the
154 remaining jars were used for reheating.

155

Reheating procedure

At every important step of the storage experiment (after sterilization and after 5 months of storage at 4 and 35 °C), a sample of each vegetable purée was taken for reheating. This sample consisted of 8 g vegetable purée which was transferred into a small glass tube (10 ml, approximately 1 cm headspace), to allow fast heating up and cooling rates, simulating the heating rates applied in microwave heating. From a worst-case perspective, the glass tubes were closed with a plastic cap. This way, a possible loss of furan during the reheating procedure was reduced as much as possible. The vegetable purées were briefly reheated by placing them in a water bath (WBU 45, Memmert, Schwabach, Germany) at 95 °C. Temperature profiles in the water bath and at the coldest point of the product were recorded using type T thermocouples (Ellab, Hilleroed, Denmark) (results not shown). When a product temperature of 90 °C was reached, the samples were removed from the water bath and immediately transferred to iced water to reduce further chemical reactions. Depending on the type of vegetable purée, this resulted in a heating time between 3 and 4 min. All the reheated vegetable purées were analyzed for (methyl)furan.

Quantitation of furan and methylfuran

Furan was analyzed with solid phase microextraction coupled to gas chromatography-mass spectrometry (SPME-GC-MS), using furan-d₄ as an internal standard. The analytical procedure is described in detail elsewhere (Palmer et al. 2014), and will only be discussed here in brief. Standard stock (ca. 2.5 mg/ml in methanol) and working solutions (ca. 0.25 µg/ml in deionized water) of internal standard were obtained by making a serial dilution of furan-d₄ (98%, Sigma Aldrich, Saint Louis, Missouri) in closed headspace vials of 10 ml. Both the amount of solvent and the amount of solution added were determined by differential weighing and the exact concentration of furan-d₄ was calculated from the respective masses. The samples were prepared for analysis by weighing 2.5 g of the cooled purée in an empty 10 ml headspace vial with a PTFE/silicone septum seal. Before the vial was completely closed, 2.5 ml of a saturated NaCl solution was added to the purée and the mixture was further diluted with deionized water to obtain a standardized total volume of 6 ml. After sealing, 100 µl

182 of the internal standard working solution was added with a chilled gastight syringe. The exact amounts
183 were determined again by differential weighing. All operations were done in a closed and refrigerated
184 sample preparation box (MPR-311D(H), Sanyo, Moriguchi, Japan) and the samples were stored at a
185 temperature of 10 °C until analysis.

186 The analyses were carried out using an Agilent 7890A GC and an Agilent 5975C MS (Keysight
187 Technologies, Santa Rosa, California), equipped with a CTC Combi PAL autosampler (CTC Analytics,
188 Zwingen, Switzerland). The SPME fiber (Supelco, Bellefonte, Pennsylvania) had a 85 µm
189 carboxen/polydimethylsiloxane (CAR/PDMS) sorptive coating, which was exposed to the headspace of
190 the samples for 15 min at 30 °C. After headspace extraction, the fiber was transferred to the GC
191 injection port, where the adsorbed compounds were thermally desorbed for 1 min at 200 °C. After each
192 run, the fiber was thermally cleaned for 2 min at 300 °C in the conditioning station of the autosampler.
193 The volatiles were injected in the splitless mode and subsequently separated on a HP-PLOT Q column
194 (30 m × 320 µm, 20 µm film thickness, Keysight Technologies, Santa Rosa, California), using helium
195 as the carrier gas at a constant flow rate of 2 ml/min. The column oven was programmed at a starting
196 temperature of 40 °C, which was retained for 4 min, after which it was elevated to 160 °C at a rate of 40
197 °C/min, followed by a second ramp to 220 °C at 5 °C/min. After 1 min at the final temperature, the oven
198 was cooled again to the initial temperature. Mass spectra were obtained by electron ionisation (EI) at 70
199 eV, in the combined SCAN and SIM mode. The scanning range extended m/z 35-400. The selected ions
200 monitored were m/z 68 (quantifier) and 39 (qualifier) for furan and m/z 72 (quantifier), 44 and 42 (both
201 qualifier) for furan-d₄. MS ion source and quadrupole temperatures were 230 and 150 °C, respectively.
202 Each sample was analyzed in duplicate.

203 The method of internal standard calibration was used to prepare a calibration curve for furan. A serial
204 dilution was made starting from a furan working solution (ca. 0.25 µg/ml in deionized water), resulting
205 in a calibration curve covering the concentration range of 0-50 ng/g purée. The identity of both furan
206 and furan-d₄ was confirmed by calculating the response ratio of the qualifier ions and the quantifier ions
207 according to the guidelines stated in European Commission Decision (European Commission 2002). In

208 agreement with these guidelines, the method was validated in terms of specificity, recovery, precision
209 (repeatability), decision limit ($CC\alpha$), and detection capability ($CC\beta$). The decision limit and the
210 detection capability of the procedure were 1.15 ng and 1.86 ng per g purée, respectively. Next to furan,
211 the calibration curve was used to estimate the concentrations of 2- and 3-methylfuran in the samples.
212 According to our own findings (results not shown), the behavior of both methylfurans towards the
213 internal standard furan- d_4 was not significantly different from the behavior of furan. The identity of 2-
214 and 3-methylfuran was confirmed by comparison with retention times of standards and by calculating
215 the response ratio of the qualifier ions and the quantifier ions, as analogous to the identification of furan
216 and furan- d_4 . For both methylfurans, the selected ions monitored were m/z 82 (quantifier), 81 and 53
217 (qualifier).

218

219 **Results and discussion**

220 Seven vegetable purées (broccoli, orange carrot, pea, potato, pumpkin, red beet and spinach) were
221 selected to investigate the effects of storage and reheating on the furan concentrations of sterilized,
222 vegetable-based foods. Some general trends on the methylfuran concentrations of these vegetable purées
223 are discussed afterwards.

224

225 ***Furan formation during storage of sterilized vegetable purées***

226 Prior to storage, the vegetable purées were subjected to a thermal treatment, aiming at an industrially
227 relevant process value for sterilization ($F_0 = 5$ min). The sterilized vegetable purées were analyzed for
228 furan and the resulting concentrations are depicted in **Fig. 1**. The range of furan concentrations found
229 after thermal treatment was very comparable to the amounts observed in our previous study (Palmer et
230 al. 2014). Red beet was again the vegetable type which was the most susceptible to furan formation (11
231 ng/g purée), followed by potato and carrot (both 7 ng/g purée). Broccoli, pea, pumpkin and tomato had a
232 comparable, lower amount of furan (3-5 ng/g purée). To gain insight into the effects of storage on the
233 furan concentrations of vegetable-based foods, the sterilized vegetable purées were stored at two

different temperatures (i.e. 4 and 35 °C). The furan concentrations of the vegetable purées stored at the latter temperature are represented in **Fig. 1**, together with the furan concentrations after sterilization. For most of the vegetable purées, the furan concentrations increased during the storage period of 5 months at 35 °C. The extent of this increase was matrix-dependent. A large increase was observed for potato (+283%) and pumpkin (+396%), in which furan concentrations were four to five times higher as compared with the concentrations observed after sterilization. A smaller increase was observed for broccoli (+114%), carrot (+35%) and tomato (+52%). Pea and red beet were the only vegetable purées in which the furan concentrations did not further increase during storage. For red beet, this was a remarkable observation, given the high furan concentration found immediately after sterilization.

With the observed increase in furan concentrations during storage, final furan concentrations up to 19 (pumpkin) and 28 ng/g purée (potato) were measured. These values were closer to the concentrations found in real vegetable-based products (mean concentrations of 23-24 ng/g for soups, 48-49 ng/g for vegetable-based baby foods) (European Food Safety Authority (EFSA) 2011) than the furan concentrations found immediately after thermal treatment. Therefore, the furan concentrations found in commercial samples can probably be explained by the summed effect of thermal processing and storage. Of course, the specific storage conditions applied for these products will determine to what extent furan forming reactions can occur during storage. The major driving force of these reactions is the storage temperature, but also water activity, pH, packaging and radiation can influence the degradation of food components during storage (Narayan 1997). To evaluate the potential of reducing furan formation during storage, the sterilized vegetable purées were stored at a refrigerated temperature of 4 °C. The storage time was 5 months, like for the vegetable purées stored at 35 °C. The resulting furan concentrations are represented in **Fig. 2**, together with the concentrations after sterilization. The results indicated that the storage temperature plays an important role in the furan formation of vegetable-based products. At 4 °C, the furan concentrations of all the selected vegetable purées remained approximately constant as compared with the values immediately after sterilization. This included potato and pumpkin purée, vegetable purées that were characterized by a large increase in furan concentrations during

260 storage at 35 °C. In other words, by selecting appropriate (refrigerated) storage conditions for sterilized,
261 vegetable-based products, the furan formation during storage could be reduced to a large extent.

262 Unlike the thermal treatment, no literature information can be found on the effects of storage for the
263 furan concentrations of vegetable-based food products. At the temperature-abuse conditions applied in
264 this study, there was a clear increase in the furan concentrations of most vegetable purées. The increase
265 in furan concentrations can be attributed to the continued degradation of precursors and intermediate
266 products of furan formation during storage. For potato and pumpkin, the extent of the increase during
267 storage was remarkably large. Since all vegetable purées were stored under the same conditions of
268 reduced oxygen and/or light exposure, it seems unlikely that these extrinsic parameters were responsible
269 for the differences in matrix susceptibility. It is possible that the sterilization treatment generated an
270 additional amount of reactive furan precursors in these matrices (e.g. by thermal degradation of
271 biomolecules such as starch and proteins), which would result in an increased furan formation during
272 storage. In order to control this furan formation in a matrix-specific way, a deeper understanding of the
273 reaction pathways leading to furan formation is indispensable, and should be subject of future research.

274

275 ***Furan formation during reheating of vegetable purées***

276 Most commercially available foods are accompanied by short guidelines on how to prepare that
277 specific product for consumption. For sterilized, vegetable-based foods, these guidelines often include a
278 reheating step. As mentioned in the introduction, reheating of the product is the final preparation step in
279 which the furan concentration is likely to be affected, before consumption. In order to elucidate the
280 general impact of this reheating step on the furan concentrations of sterilized, vegetable-based food
281 products, the thermally treated and stored vegetable purées of the previous section were briefly reheated
282 to 90 °C (3-4 min, depending on the vegetable type). After the reheating step, the vegetable purées were
283 analyzed for furan. In **Table 1**, the results of these analyses are compared with the concentrations of
284 furan after storage, which were already discussed in Section 3.1. In general, furan concentrations
285 decreased as a result of the reheating step. The largest decrease in furan concentrations was observed for

286 potato (-44%) and pumpkin purée (-70%) stored at 35 °C, which were also the vegetable purées with the
287 highest furan concentrations after storage. When the vegetable purées were characterized by lower
288 initial amounts of furan, the reheating procedure seemed to have a smaller influence on the furan
289 concentrations. As a result, many vegetable purées showed only a small decrease in terms of absolute
290 values (0-2 ng/g purée). Broccoli and tomato purées were almost unaffected by reheating.

291 Theoretically, a reheating step can influence the furan concentrations of sterilized, vegetable-based
292 food products in two ways (Hasnip et al. 2006; Roberts et al. 2008; Lachenmeier et al. 2009). By
293 heating up the product, both the mobility and the volatility of furan are increased, which can result in
294 evaporative losses of furan to the surroundings. At the same time, an additional amount of furan can be
295 formed due to thermal degradation reactions inside the product. The overall decrease in furan
296 concentrations indicated that for this particular case, evaporation of furan was the dominating event.
297 This was not really a surprise, because the temperature-time conditions applied during the reheating
298 procedure were only a fraction of the heating conditions applied during sterilization. Unless the product
299 is (accidentally) exposed to high temperatures or long heating times, the reheating step is not expected to
300 cause a significant increase of the furan concentration in the product. The observed decrease in furan
301 concentrations after reheating of the sterilized or stored vegetable purées corresponds to results reported
302 in the literature. Zoller et al. (2007), Roberts et al. (2008) and Kim et al. (2009) have all demonstrated
303 furan concentrations to be reduced to roughly half the initial concentration after applying different
304 reheating procedures (open/closed heating, microwave/saucepan heating, stirring) to commercially
305 available, ready-to-eat foods. The maximum heating temperature seems to play an important role for the
306 extent of this decrease (Fromberg et al. 2009). When milder procedures of reheating were applied, furan
307 was found to be more persistent (Hasnip et al. 2006; Lachenmeier et al. 2009). The retention of furan in
308 ready-to-eat foods can also be influenced by the presence of lipophilic compounds (such as oils) in the
309 product (Van Lancker et al. 2009), which is an important difference with the vegetable purées used in
310 the present study (no oil/fat added). Furthermore, it is possible that the structural properties of the

311 vegetable purées affect the extent of the furan evaporation (Van Lancker et al. 2009), but this was not
312 clearly observed in the present study.

313

314 *Trends on the methylfuran concentrations of vegetable purées*

315 Next to furan, the stored and reheated vegetable purées were also analyzed for 2- and 3-methylfuran.
316 Due to the little amount of literature data on the toxicological relevance and the reaction pathways
317 leading to the formation of both methylfurans, the discussion will be limited to a qualitative description
318 of the changes in methylfuran concentrations as influenced by both effects. For most vegetables, the
319 concentrations of both alkylated furan derivatives were very similar to each other. To facilitate their
320 discussion, the concentrations were summed together as a group of methylfuran compounds.

321 The summed methylfuran concentrations of the vegetable purées after sterilization and storage at 35
322 °C are represented in **Fig. 3**. Immediately after the thermal treatment for sterilization, very low amounts
323 of summed methylfuran (1-3 ng/g purée) were found in most vegetable purées. Tomato and carrot purée
324 were the exceptions, with a concentration of 127 and 12 ng/g purée, respectively. A high susceptibility
325 to methylfuran formation for tomato-based products has been observed before (Becalski et al. 2010).
326 For this reason, tomato-based products might be considered as priority matrices with regard to
327 methylfuran mitigation, if deemed necessary. Following storage at 35 °C for 5 months, the
328 concentrations of summed methylfuran in both tomato (-90%) and carrot purée (-88%) decreased, as
329 opposed to the trends observed for furan. The low initial amounts of methylfuran in the other vegetable
330 purées were almost unaffected by the same storage conditions. Similar results were observed for the
331 vegetable purées stored at 4 °C (results not shown), although the decrease in methylfuran concentrations
332 for tomato (-24%) and carrot purée (-55%) was clearly smaller at this lower storage temperature. The
333 decrease in methylfuran concentrations at both storage temperatures is markedly different from the
334 observed increase for furan. 2- and 3-methylfuran seemed to be prone to degradation reactions during
335 storage, whereas furan was considered to be a rather stable molecule based on the results of the storage
336 experiment described above (cf. Section 3.1). Together with the clearly different matrix dependency of

337 methylfuran formation during thermal treatment for sterilization, these results seem to indicate different
338 reaction pathways for both types of compounds, which would further complicate the mitigation of both
339 compounds in thermally treated foods.

340 The majority of the vegetable purées tested in the present study showed a (small) decrease in summed
341 methylfuran concentrations after the reheating step prior to consumption (results not shown). Like for
342 furan, the decrease in methylfuran concentrations could most probably be explained by evaporative
343 losses during the reheating procedure and/or partial degradation of both compounds as observed during
344 storage. The decrease in methylfuran concentrations due to the reheating procedure was less pronounced
345 than for furan. Most vegetable purées already had low initial amounts of methylfuran before reheating,
346 and moreover, 2- and 3-methylfuran molecules are less volatile (boiling point around 63-66 °C) as
347 compared with furan (31 °C). With an optimized procedure of reheating, it should be possible to
348 simultaneously reduce furan and methylfuran concentrations of thermally treated foods.

349

350 **Conclusions**

351 The present study has demonstrated possible furan formation during storage of vegetable-based foods.
352 Following storage for 5 months at 35 °C (temperature-abuse conditions), the furan concentrations of a
353 range of sterilized vegetable purées were increased in a matrix-specific manner. The storage temperature
354 had an important impact on the extent of furan formation during storage. At a refrigerated storage
355 temperature of 4 °C, the furan concentrations of the vegetable purées remained approximately constant
356 for the same period of 5 months. Even though storage at refrigerated temperatures will contribute to the
357 general quality stability of the product, this mitigation strategy might not be a realistic option from an
358 economic point of view. Therefore, research should continue on the elucidation of the relationship
359 between storage conditions and matrix composition on the one side and furan formation on the other
360 side (e.g. by performing kinetic or mechanistic studies). In particular, future experiments should target
361 possible furan formation at room temperatures, which are the typical storage temperatures of sterilized,
362 vegetable-based foods. With regard to the effect of the matrix composition, there are a multitude of

factors that can possibly explain the observed matrix-specificity during storage (e.g. precursors, redox condition, pH). A better understanding of the reaction pathways leading to furan, is a logical next step towards control of furan formation in critical matrices, such as the potato and pumpkin purées in the present study. In the search for effective furan mitigation measures that do not compromise general food safety nor quality, the effect of the reheating step prior to consumption was also investigated. As contrary to storage, furan concentrations decreased when the sterilized and stored vegetable purées were subjected to a short reheating step. At the temperature-time conditions applied in this study (3-4 min of reheating to 90 °C), furan was most likely lost due to evaporative losses. Many additional parameters can influence the extent of this evaporation from the product. If further optimized and properly communicated to the general audience, the reheating procedure could result in an easy and reliable way for removing part of the furan concentration in critical products, such as jarred baby foods. In view of their possible toxicological relevance to the risk characterization of furan, the vegetable purées were also analyzed for 2- and 3-methylfuran. Both methylfurans were mainly formed during the thermal treatment for sterilization. Tomato purée was found to be very susceptible to the formation of methylfuran, as opposed to most of the other vegetable purées. An overall decrease in the methylfuran concentrations was observed during storage, which was contrary to the results observed for furan. The concentrations further decreased after reheating of the purées, which could be explained by evaporative losses and/or partial degradation of 2- and 3-methylfuran.

Acknowledgements

Stijn Palmers is funded by the Agency for Innovation by Science and Technology in Flanders (IWT), co-authors Tara Grauwet and Biniam T. Kebede by the Research Foundation Flanders (FWO) and the KU Leuven Research Fund, respectively.

387 **References**

- 388 Adebooye OC, Vijayalakshmi R, Singh V. 2008. Peroxidase activity, chlorophylls and antioxidant
389 profile of two leaf vegetables (*Solanum nigrum* L. and *Amaranthus cruentus* L.) under six
390 pretreatment methods before cooking. *Int J Food Sci Technol*. 43:173-178.
- 391 Becalski A, Hayward S, Krakalovich T, Pelletier L, Roscoe V, Vavasour E. 2010. Development of an
392 analytical method and survey of foods for furan, 2-methylfuran and 3-methylfuran with
393 estimated exposure. *Food Additives and Contaminants Part A-Chemistry Analysis Control*
394 *Exposure & Risk Assessment*. 27:764-775.
- 395 Becalski A, Seaman S. 2005. Furan precursors in food: A model study and development of a simple
396 headspace method for determination of furan. *Journal of Aoac International*. 88:102-106.
- 397 European Commission. 2002. Commission Decision of 12 August 2002 implementing Council
398 Directive 96/23/EC concerning the performance of analytical methods and the interpretation of
399 results. 2002/657/EC:
- 400 European Food Safety Authority (EFSA). 2011. Update on furan levels in food from monitoring years
401 2004-2010 and exposure assessment. *EFSA Journal*. 9:1-33.
- 402 Fan XT. 2005. Formation of furan from carbohydrates and ascorbic acid following exposure to ionizing
403 radiation and thermal processing. *J Agric Food Chem*. 53:7826-7831.
- 404 Fromberg A, Fagt S, Granby K. 2009. Furan in heat processed food products including home cooked
405 food products and ready-to-eat foods. European Food Safety Authority (EFSA). EFSA-Q-2009-
406 00846.
- 407 Gill S, Kavanagh M, Cherry W, Barker M, Weld M, Cooke GM. 2014a. A 28-day Gavage Toxicity
408 Study in Fischer 344 Rats with 3-methylfuran. *Toxicologic Pathology*. DOI:
409 10.1177/0192623314534537.
- 410 Gill SS, Kavanagh M, Cherry W, Barker M, Weld M, Cooke GM. 2014b. A 28-day Gavage Toxicity
411 Study in Male Fischer 344 Rats with 2-methylfuran. *Toxicologic Pathology*. 42:352-360.
- 412 Hasnip S, Crews C, Castle L. 2006. Some factors affecting the formation of furan in heated foods. *Food*
413 *Additives and Contaminants*. 23:219-227.
- 414 Huang XS, Duan HY, Barringer SA. 2011. Effects of buffer and temperature on formation of furan,
415 acetic acid and formic acid from carbohydrate model systems. *Lwt-Food Science and*
416 *Technology*. 44:1761-1765.
- 417 International Agency for Research on Cancer (IARC). 1995. Dry cleaning, some chlorinated solvents
418 and other industrial chemicals. IARC Monographs on the Evaluation of Carcinogenic Risks to
419 Humans. 63:393-407.
- 420 Joint FAO/WHO Codex Committee on Contaminants in Foods. 2011. Discussion paper on furan
421 (CX/CF 11/5/13).
- 422 Kim TK, Lee YK, Park YS, Lee KG. 2009. Effect of cooking or handling conditions on the furan levels
423 of processed foods. *Food Additives and Contaminants Part A-Chemistry Analysis Control*
424 *Exposure & Risk Assessment*. 26:767-775.

425 Lachenmeier DW, Reusch H, Kuballa T. 2009. Risk assessment of furan in commercially jarred baby
426 foods, including insights into its occurrence and formation in freshly home-cooked foods for
427 infants and young children. *Food Additives and Contaminants Part A-Chemistry Analysis*
428 *Control Exposure & Risk Assessment*. 26:776-785.

429 Limacher A, Kerler J, Conde-Petit B, Blank I. 2007. Formation of furan and methylfuran from ascorbic
430 acid in model systems and food. *Food Additives and Contaminants*. 24:122-135.

431 Limacher A, Kerler J, Davidek T, Schmalzried F, Blank I. 2008. Formation of furan and methylfuran by
432 Maillard-type reactions in model systems and food. *J Agric Food Chem*. 56:3639-3647.

433 Locas CP, Yaylayan VA. 2004. Origin and mechanistic pathways of formation of the parent furan - A
434 food toxicant. *J Agric Food Chem*. 52:6830-6836.

435 Mark J, Pollien P, Lindinger C, Blank I, Mark T. 2006. Quantitation of furan and methylfuran formed in
436 different precursor systems by proton transfer reaction mass spectrometry. *J Agric Food Chem*.
437 54:2786-2793.

438 Narayan KA. 1997. *Food Storage Stability*. CRC Press. Biochemical aspects;

439 Owczarek-Fendor A, De Meulenaer B, Scholl G, Adams A, Van Lancker F, Eppe G, De Pauw E,
440 Scippo ML, De Kimpe N. 2012. Furan formation in starch-based model systems containing
441 carbohydrates in combination with proteins, ascorbic acid and lipids. *Food Chemistry*. 133:816-
442 821.

443 Owczarek-Fendor A, De Meulenaer B, Scholl G, Adams A, Van Lancker F, Eppe G, De Pauw E,
444 Scippo ML, De Kimpe N. 2011. Furan formation from lipids in starch-based model systems, as
445 influenced by interactions with antioxidants and proteins. *J Agric Food Chem*. 59:2368-2376.

446 Owczarek-Fendor A, De Meulenaer B, Scholl G, Adams A, Van Lancker F, Yogendrarajah P, Eppe G,
447 De Pauw E, Scippo ML, De Kimpe N. 2010. Furan formation from vitamin C in a starch-based
448 model system: Influence of the reaction conditions. *Food Chemistry*. 121:1163-1170.

449 Palmers S, Grauwet T, Kebede BT, Hendrickx ME, Van Loey A. 2014. Reduction of furan formation by
450 high-pressure high-temperature treatment of individual vegetable purées. *Food Bioprocess*
451 *Technol*. 7:2679-2693.

452 Roberts D, Crews C, Grundy H, Mills C, Matthews W. 2008. Effect of consumer cooking on furan in
453 convenience foods. *Food Additives and Contaminants*. 25:25-31.

454 Taub IA, Singh RP. 1997. *Food Storage Stability*. Taylor & Francis.

455 Toledo RT. 2007. *Fundamentals of food process engineering*. Springer US. Chapter 8, Kinetics of
456 chemical reactions in foods; 285-299.

457 US Food and Drug Administration (FDA). 2009. Exploratory data on furan in food: individual food
458 products.

459 Van Lancker F, Adams A, Owczarek A, De Meulenaer B, De Kimpe N. 2009. Impact of various food
460 ingredients on the retention of furan in foods. *Molecular Nutrition & Food Research*. 53:1505-
461 1511.

462 Van Lancker F, Adams A, Owczarek-Fendor A, De Meulenaer B, De Kimpe N. 2011. Mechanistic
463 Insights into Furan Formation in Maillard Model Systems. *J Agric Food Chem.* 59:229-235.

464 Vervoort L, Van der Plancken L, Grauwet T, Verlinde P, Matser A, Hendrickx M, Van Loey A. 2012.
465 Thermal versus high pressure processing of carrots: A comparative pilot-scale study on
466 equivalent basis. *Innovative Food Science & Emerging Technologies.* 15:1-13.

467 Zoller O, Sager F, Reinhard H. 2007. Furan in food: Headspace method and product survey. *Food*
468 *Additives and Contaminants.* 24:91-107.

469

470

471

472 **List of tables**

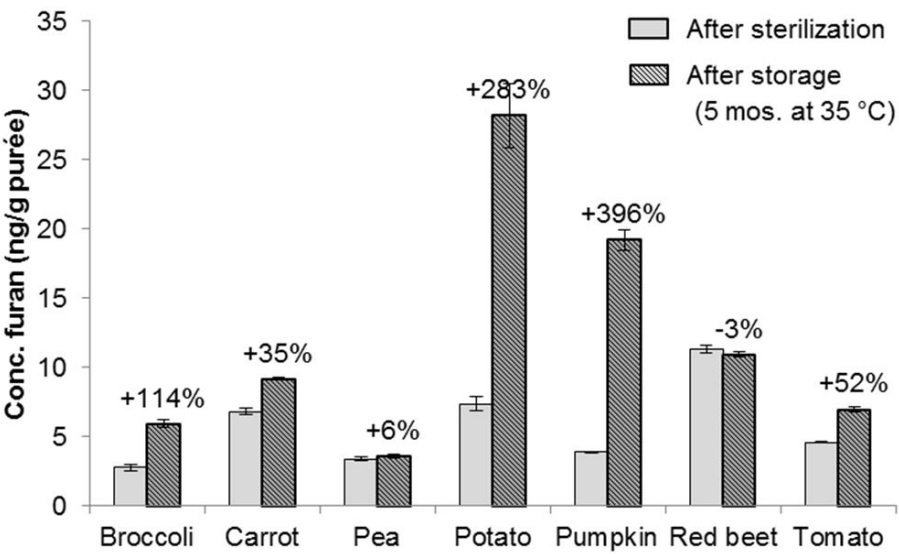
473

474 **Table 1.** Change in furan concentrations after reheating of the vegetable purées. For each vegetable
475 purée, a part of the purée after sterilization and the purées stored at 4 and 35 °C was reheated.

Matrix	Treatment	Initial furan conc. (ng/g purée)	Furan conc. after reheating (ng/g purée)	Change (±ng/g purée)	Change (%)
Broccoli	Sterilized (not stored)	2.76	3.38	+0.62	+22
	Stored at 4 °C (5 mos.)	4.18	4.08	-0.10	-2
	Stored at 35 °C (5 mos.)	5.93	5.14	-0.79	-13
Carrot	Sterilized (not stored)	6.81	3.68	-3.14	-46
	Stored at 4 °C (5 mos.)	4.88	4.38	-0.50	-10
	Stored at 35 °C (5 mos.)	9.17	7.39	-1.77	-19
Pea	Sterilized (not stored)	3.38	1.01	-2.37	-70
	Stored at 4 °C (5 mos.)	3.57	3.07	-0.50	-14
	Stored at 35 °C (5 mos.)	3.57	1.11	-2.47	-69

Potato	Sterilized (not stored)	7.36	5.47	-1.90	-26
	Stored at 4 °C (5 mos.)	7.13	3.65	-3.49	-49
	Stored at 35 °C (5 mos.)	28.22	15.75	-12.47	-44
Pumpkin	Sterilized (not stored)	3.88	1.21	-2.67	-69
	Stored at 4 °C (5 mos.)	4.76	3.98	-0.78	-16
	Stored at 35 °C (5 mos.)	19.21	5.61	-13.60	-70
Red beet	Sterilized (not stored)	11.29	9.87	-1.42	-13
	Stored at 4 °C (5 mos.)	11.89	8.63	-3.26	-27
	Stored at 35 °C (5 mos.)	10.93	8.23	-2.70	-25
Tomato	Sterilized (not stored)	4.58	5.60	+1.02	+22
	Stored at 4 °C (5 mos.)	6.39	5.96	-0.43	-7
	Stored at 35 °C (5 mos.)	6.96	6.35	-0.61	-9

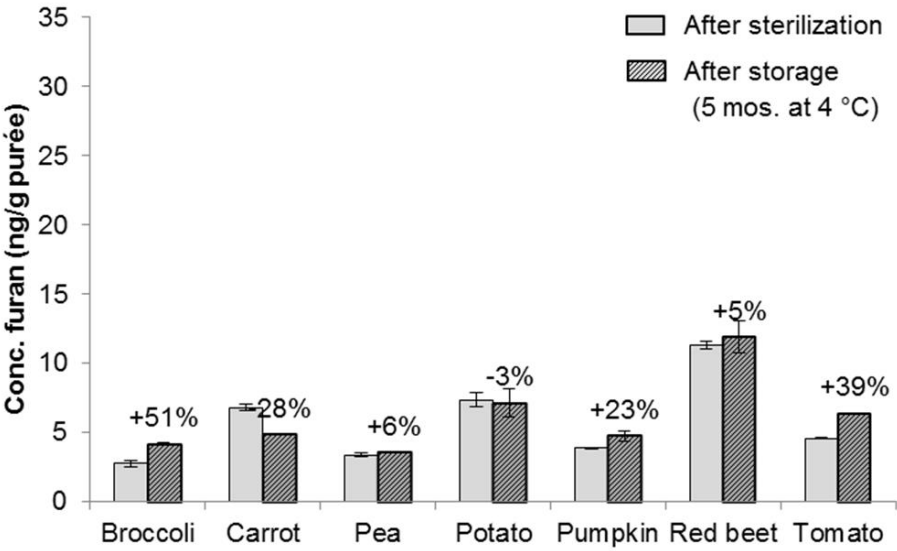
478



479

480 **Fig. 1.** Furan concentrations of the vegetable purées after sterilization (F0 = 5 min) and after 5 months
481 of storage at 35 °C. Data labels indicate the change in furan concentration (in percentages) between both
482 times of measurement.

483



485

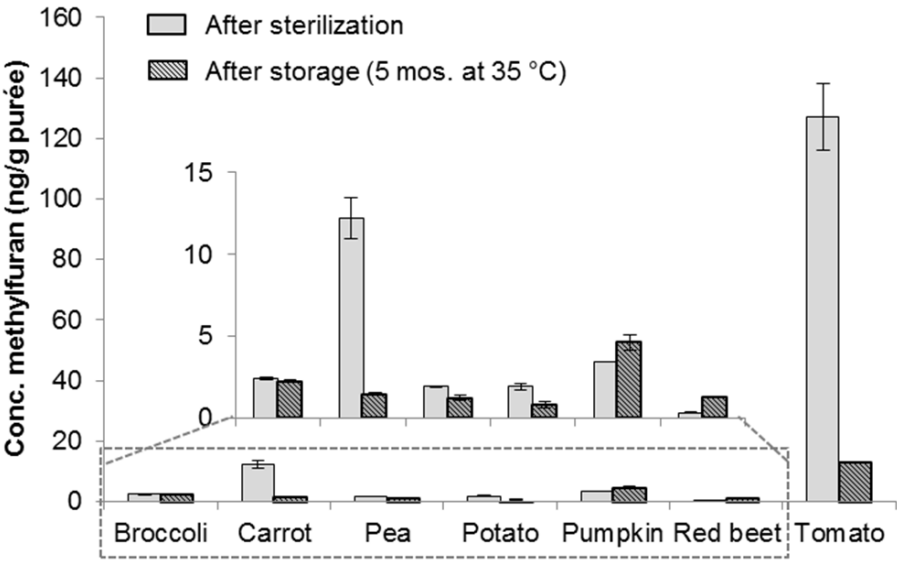
486

487

488

489

Fig. 2. Furan concentrations of the vegetable purées after sterilization ($F_0 = 5$ min) and after 5 months of storage at 4 °C. Data labels indicate the change in furan concentration (in percentages) between both times of measurement.



491

492

493

494

Fig. 3. Summed methylfuran concentrations of the vegetable purées after sterilization ($F_0 = 5$ min) and after 5 months of storage at 35 °C.